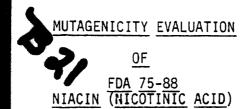


BIONETICS



FINAL REPORT

5516 Nicholson Land Kensington, Maryland 20795

MUTAGENICITY EVALUATION

<u>OF</u>

NIACIN (NICOTINIC ACID)

FINAL REPORT

SUBMITTED TO

GENETIC TOXICOLOGY BRANCH
DIVISION OF TOXICOLOGY
BUREAU OF FOODS
U.S. FOOD AND DRUG ADMINISTRATION
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EVALUATION SUMMARY

The test compound, FDA 75-88, Niacin (nicotinic acid), did not exhibit mutagenic activity in any of the assays employed in these studies.



DATE:

July, 1977

SPONSOR:

U.S. Food and Drug Administration

SUBJECT: Evaluation of Test Compound: FDA 75-88, Nitacin (nicotinic acid)

I. **OBJECTIVE**

The objective of this study was to evaluate the test compound for genetic activity in microbial assays with and without the addition of mammalian metabolic activation preparations.

II. MATERIALS

A. Test Compound

1. Date Received:

December 29, 1976

2.

Description:

White powder

В. Indicator Microorganisms

The following strains of indicator microorganisms were used in the evaluation:

Yeast Strain:

Saccharomyces cerevisiae, strain D4

Bacteria Strains:

Salmonella typhimurium, strains TA-1535

TA-1537

TA-1538

TA-98

TA-100

C. Reaction Mixture

The following reaction mixture was employed in the activation tests:

Final Concentration/ml Component 1. TPN (sodium salt) umoles 2. Glucose-6-phosphate 5 umoles 3. Sodium phosphate (dibasic) 100 umoles 4. MgCl₂ 8 umoles 5. KC1 33 µmoles 6. Homogenate fraction equivalent to 25 mg of wet tissue.



D. Tissue Homogenates and Supernatants

The tissue homogenates and 9,000 x g supernatants were prepared from tissues of the following mammalian species: Mouse - ICR random bred adult males; rat - Sprague-Dawley adult males; and monkey - Macaca mulatta adult males.

E. Positive Control Compounds

Table 1 lists chemicals for positive controls in the direct and activation assays.

TABLE 1

POSITIVE CONTROLS USED IN DIRECT AND ACTIVATION ASSAYS

Assay	<u>Chemical^a</u>	Solvent	Probable Mutagenic Specificity
Nonactivation	Methylnitrosoguanidine Ethylmethanesulfonate 2-Nitrofluorene Quinacrine mustard	Water or saline Water or saline Dimethylsulfoxide ^C Water or saline	BPSb BPSb FSb FSb
Activation	Dimethylnitrosamine 2-Acetylaminofluorene 8-Aminoquinoline 2-Aminoanthracene	Water or saline Dimethylsulfoxide Dimethylsulfoxide Dimethylsulfoxide	BPS ^b FS ^b FS ^b BPS ^b

Concentrations given in the Results Section
BPS = base-pair substitution; FS = frameshift
Previously shown to be non-mutagenic

III. METHODS

A. <u>Toxicity</u>

The solubility, toxicity and doses for the test chemical were determined prior to screening.

The test chemical was tested for toxicity against specific indicator strains over a range of doses to determine the 50% survival dose. Bacteria were tested in phosphate buffer, pH 7.4, for one hour at 37°C on a shaker. Yeasts were tested in phosphate buffer, pH 7.4, for four hours at 30°C on a shaker. The 50% survival concentrations and the 1/4 and 1/2 50% doses calculated.

If no toxicity was obtained for the chemical with a given strain, then a maximum dose of 5% (w/v) was used.

Unless otherwise specified, the doses calculated for the tests in buffer were applied to the activation tests. The solubility of the test chemical under treatment conditions is stated in the Results Section.



B. Plate Tests (Overlay Method)

Approximately 10⁸ cells from an overnight culture of each indicator strain were added to test tubes containing 2.0 ml of molten agar supplemented with biotin and a trace of histidine. For nonactivation tests, the three dose levels of the test compound were added to the contents of the appropriate tubes and poured over the surfaces of selective agar plates. In activation tests 0.5 ml of a 9,000 x g tissue supernatant and required cofactors (core reaction mixture) were added to the overlay tubes. Three dose levels of the test chemical were added to the appropriate tubes, which were then mixed and the contents poured over the surface of a minimal agar (selective medium) plate and allowed to solidify. The plates were incubated for 48 to 72 hours at 37°C, and scored for the number of colonies growing on each plate. The concentrations of all chemicals are given in the Results Section. Positive and solvent controls using positive compounds that are active directly and those that require metabolic activation were run with each assay.

C. <u>Suspension Tests</u>

Nonactivation

Bacteria and yeast cultures of the indicator organisms were grown in complete broth, washed and resuspended in 0.9% saline to densities of 1 \times 10¹⁰ cells/ml and 5 x 10° cells/ml, respectively. This constituted the working stock for tests of a group of test chemicals and their respective controls. Tests were conducted in plastic, 24-well tissue culture plates (Linbro). Cells plus appropriate volume(s) of the test chemical were added to the wells to give a final volume of 1.5 ml. The solvent replaced the test chemical in the negative controls. Treatment was at 30°C for four hours for yeast tests and at 37°C for one hour for bacterial tests. All flasks were shaken during treatment. Following treatment, the plates were set on ice. Aliquots of cells were removed, diluted in sterile saline (4°C) and plated on the appropriate complete media. Undiluted samples from flasks containing the bacteria were plated on minimal selective medium in reversion experiments. Samples from a 10 1 dilution of treated cells were plated on the selected media for enumeration of gene conversion with strain D4. Bacterial plates were scored after incubation for 48 hours at 37°C. The yeast plates were incubated at 30°C for 3-5 days before scoring.

2. Activation

Bacteria and yeast cells were grown and prepared as described in the nonactivation tests. Measured amounts of the test and control chemicals plus 0.25 ml of the stock-cell suspension were added to wells of the Linbro plate containing the appropriate tissue fraction and reaction mixture. All flasks (bacteria and yeast) were incubated at 37°C with shaking. The treatment times as well as the dilutions, plating procedures and scoring of the plates were the same as described for nonactivation tests.



D. Preparation of Tissue Homogenates and 9,000 x g Cell Fractions

Male animals (except monkeys) sufficient to provide the necessary quantities of tissues were killed by cranial blow, decapitated and bled. Monkey tissues were obtained from freshly killed and bled male rhesus monkeys. Organs were immediately dissected from the animals using aseptic techniques and placed in ice-cold 0.15M KCl. Upon collection of the desired quantity of organs, they were washed twice with fresh KCl and completely homogenized with a motor-driven homogenizing unit at 4° C. The whole organ homogenate obtained from this step was divided into two samples. One sample was frozen at -80° C and the other was centrifuged for 20 minutes at 9,000 x g in a refrigerated centrifuge, The supernatant from the centrifuged sample was retained and frozen at -80° C. These two frozen samples were used for the activation studies. Protein and P-448 determinations were made for each lot of homogenate.

E. Data Recording and Reporting

1. Plate test assays

The numbers of colonies on each plate were counted and recorded on printed forms. These raw data were entered into a computer program designed to print out all data by test. The data are presented as revertants per plate for each indicator strain employed in the assay. The positive and solvent controls are provided as reference points.

2. Suspension assays

Following the specified incubation periods all population plates were scored by an automatic colony counter and the results from each plate of a set were recorded, in ink, on data processing forms. All minimal or other types of selective media plates were hand scored and the results recorded along with the respective population data. Other relevant experimental data were recorded on experimental definition forms. For bacteria strains the number of colonies recorded from either the population or selective plates represents that number in 1 ml of test suspension plated. The numbers recorded for the yeast strain D4 represent the number in 0.5 ml of test suspension plated. The data were then processed and printed from a computer program. All raw data sheets are dated and signed by the responsible technician.



- IV. RESULTS SECTION
- A. Solubility Properties of the Test Compound
- 1. Name or code designation of the test compound: FDA 75-88, Niacin (nicotinic acid)
- 2. Test solvent: * Saline
- 3. Solubility of the test compound under treatment conditions: Soluble
- 4. Additional comments: White powder
- B. Toxicity and Dosage Determinations for the Test Compound
- 1. Test date for toxicity determination: April 4, 1977
- 2. The 50% survival level was determined for bacteria and yeast indicator organisms by conducting survival curves with the test compound at the following concentrations:

Percent Concentration (w/v or v/v)

5.0

0.5

0.05

0.005

0.0005

3. Concentrations of the test compound used in the mutagenicity tests:

	Percent Concentration					
Test Doses	Bacteria	Yeast				
1/4 50% Survival	0.004	0.0825				
1/2 50% Survival	0.008	0.1650				
50% Survival	0.016	0.3300				

^{*}The concentration of solvent was equal to the highest volume of test material added.



C. Plate Test Results

The plate test results are summarized in the following table. The values presented in this table are the number of revertants per plate.

D. Suspension Assay Results

The suspension test results for the test compound are summarized in the tables following the plate test summary. The values presented in these tables are the calculated mutation frequencies for each control and experimental test point. The first table of the suspension set presents the results for the nonactivation assays, and the second table through the fourth table of the suspension set presents the results for the activation assays. A listing of computer codes and abbreviations is included for reference. Tabulation of all raw data is provided in the Appendix.



SUMMARY OF TEST RESULTS

PLAIE_IESIS

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: 000059676

. TEST DATE: HAY 18, 1977

								E.B.I	AHI	<u>S_P</u>	F.R	PLA	<u> </u>		
IES	I			SPECIES	LISSUE	IA:	:1535_	IA	-1537_	IA	-1538_	_IA:	-98	_IA=	
	_					1	2	1	2	1	2	1	2	1	2
1.	HOM:ACII														
	SOLVENT					28	21	22	35	17	16	34	27	140	143
	POSITIVE	CONTROL				>1000	>1000	>1000	>1000	>1000	>1000	>1000	>1000	>1000	>1000
	TEST	0.0.016	\$			26	13	19	21	13	12	23	24	143	145
		4-0.008	*			14	18	17	10	10	10	30	32	146	124
		9 - 0.004	\$			15	30	19	10	12	7	27	31	129	131
2.	ACILYAII	ON													
	SOLVENT	CONTROL	•	HOUSE	LIVER	30	31	22	23	19	10	37	32	222	195
				RAT	LIVER	26	37	28	18	19	17	39	40	147	102
				MONKEY	LIVER	18	15	17	31	23	21	36	37	192	133
	POSITIVE	CONTROL	***	HOUSE	LIVER	502	498	268	256	874	911	>1000	>1000	424	889
				RAT	LIVER	274	374	241	149	938	732			>1800	>1000
				NGNKEY	LIVER	370	215	173	160	738	901	>1000	937	>1000	>1000
	TEST	0.016	. \$	HOUSE	LIVER	14	20	20	26	12	18	36	29	114	106
		0.008	4	MOUSE	LIVER	16	13	20	22	16	19	32	31	98	100
		0.004	\$	NOUSE	LIVER	14	11	12	12	13	11	32	31	117	115
		0.016	\$	RAT	LIVER	16	15	19	26	16	11	35	34	124	122
		0.008	1	RAT	LIVER	23	26	21	23	20	13	24	29	133	139
		0.004	\$	RAT	LIVER	18	15	14	15	17	20	30	26	188	141
		0.016	Š	MONKEY	LIVER	15	16	21	22	18	14	38	29	151	150
		0.008	Ē	MONKEY	LIVER	21	33	22	15	12	10	37	36	148	152
		0.004	<u>.</u>	HONKEY	LIVER	21	23	14	17	18	22	37	39	148	129
		J. 55 1	-	MANUEL S	e taru	£ £	23			10	22	31	37	170	167

NON-ACTIVATION ASSAYS CONSIST OF THE CELLS PLUS THE TEST COMPOUND VEHICLE (SOLVENT). FOR ACTIVATION ASSAYS, THE OVERLAY CONTAINS THE ACTIVATION SYSTEM PLUS THE TEST COMPOUND VEHICLE.

**	TA-1535	MNNG	2 (JG/PLATE				TA-1535	ANTH	100	UG/PLATE	
	TA-1537	QM	20 (JG/PLATE	:			TA-1537	ANG	100	UG/PLATE	
	TA-1538	NF	100	JG/PLATE	:			TA-1538	AAF	100	UG/PLATE	
	TA-98	NF	100 1	US/PLATE	:			TA-98	AAF	100	UG/PLATE	
	TA-100	HNNG	2 1	JG/PLATE				TA-100	ANTH	100	UG/PLATE	
	NOTE:	CONCENT	RATIO	ONS ARE	GIVEN	IN	HICROLITE	ERS (UL)	OR MICRO	GRAH	S(UG) PER	PLATE.

α

COMPOUND FREQUENCY SUMMARY REPORT 07/22/77

NONACTIVATION COMPOUND 000059676

TEST	ORG	TAIDO HIS EX-8	TA1535 HIS EX-8	TA1537 H15 EX-8	TA1538 HIS EX-8	TA98 HIS EX-8	ADE EX-5	FRY EX-5					
NAN		85.71	3-58	11.59	5.46	13.63	19.91	6.89	CONTROLS				
NAP		900.49	685.45	235.32	163.30	71.70	109.36	78.06					
NAI		70.81	3.58	6.89	3.52	10.35	18.61	0.06	TEST DATA				
SAM		54.17	2.42	2.87	3.86	7.66	15.07	4.78		•			
MAG		72 50	2 14	9.71	2 30	8 40	10 14	E 33			•	•	

COMPOUND FREQUENCY SUMMARY REPORT 08/04/77

SPECIES ICRFLO/MOUSE

COMPOUND 000059676

TEST	ORG	TALOO HIS EX-R	TA1535 HIS EX-8	TA1537 HIS EX-8	TA1537 HIS EX-8	TA1538 HIS EX-8	TA98 HIS EX-8	000004 ADE EX-5	0000D4 TRY EX-5	
ACT	A+C	84.43	7.09	9.31		9.43	9.46	6.93	15.63	NEGATIVE CONTROLS
ACT	A-C	55.84	4.18	6.29		8.48	8.79	6.90	22.67	
ACT	ALI	57.41	5.86	12.74	5.77	5.51	21.48	10.38	6.14	
ACT	ALU									
ACT	PLI	182.07		97.65			87.28		91.82	POSITIVE CONTROLS
ACT	PĽU	-					74.60		17.34	
ACT	LII				2.90				9.68	TEST COMPOUND
ACT	r15	74.18	3.20	10.24	2.44	7.11	18.43	1.20	0.50	
ACT	LI3	68.56	2.65	5.65	2.88	8.08	15.02	5.66	1.72	
ACT	FOI	24.82	3.53	47.21	2.64	9.54	16.71	1.85	1.12	
ACT	rns	28.39	3.28	17.79	3.50	6.41	13.41	1.59	1.20	
ACT	LU3	30.05	3.71	26.67	2.22	9.24	12.41	6.31	5.58	

COMPOUND FREQUENCY SUMMARY REPORT 07/22/77

SPECIES SPRDAW/RAT

COMPOUND 999959676

TEST	ORG	TA100 HIS EX-8	TA1535 HIS EX-8	TA1537 HIS B-X3	TA1538 HIS EX-8	HIS HIS EX-B	0000D4 ADE EX-5	0000D4 TRY EX-5	
ACT	A+C	20.79	1.37	2.13	10.56	13.11	15.60	10.18	NEGATIVE CONTROLS
ACT	A-C	18.64	3.80	6.21	5.06	15.56	12.85	7.44	
ACT	ALI	83.48	2.64	7.26	10.82	36.48	15.01	10.34	
ACT	ALU		1.76	2.62		17.63	15.67	10.27	
ACT						297.10	110.32	71.61	POSITIVE CONTROLS
ACT	PLU	52.63	1.29	3.16	164.05	124.87	17.35	7.69	
ACT	LII	16.37	3.31	7.65	19.95	35.54	11.27	12.51	TEST COMPOUND
ACT	L12	31.22	2.93	8.09	6.33	31.91	14.78	11.65	
ACT	LI3	9.57	4.65	7.62	4.52	42.49	12.95	5.25	
ACT	LUI	53.35	1.78	7.00	10.77	34.70	10.43	3,84	
ACT	LU2	46.00	1.98	6.81	7.00	16.83	8.48	4.32	
ACT	LU3	46.06	2.19	4.24	6.37	22.56	11.57	4.79	

COMPOUND FREQUENCY SUMMARY REPORT 07/22/77

SPECIES RHESUS/HONKEY

CUMPOUND 898859676

TEST	ORG	TA100 HIS B-X3	TA1535 HIS EX-8	TA1537 HIS EX-8	TA1538 HIS EX-8	HIS HIS 8-X3	0000D4 ADE EX-5	0000D4 TRY EX-5	
ACT	A+C	87.84	5.64	21.88	13.86	7.51	21.98	10.90	NEGATIVE CONTROLS
ACT	A-C	64.54	2.96	1.25	11.47	8.19	8.94	7,30	
ACT	ALI	86.09	5.54	4.98	12.37	14.55	16.25	6.21	
ACT	ALU	70.23	3.78	7.32	10.16	10.45	20.74	7.73	
ACT	PLI	284.60	56.90	37.74	154.01	201.65	68.25	54.23	POSITIVE CONTROLS
ACT	PLU	69.20	4.38	13.64	7.74	8.22	20.46	7.34	
ACT	LII	64.71	5.30	6.41	20.86	12.67	4.07	1.08	TEST COMPOUND
ACT	F15	69.81	6.31	4.81	12.85	12.70	4.88	1.73	
ACT	L13	62.35	4.00	2.21	14.65	12.39	4.94	2.19	
ACT	F01	75.42	4.40	4,45	21.86	17.73	4.27	2.40	•
ACT	LUZ	62.22	4.04	4.61	9.57	11.59	7.22	2.28	
ACT	LU3	85.91	4.10	3.77	13.92	11.55	7.11	2.65	

DATA TABLE TERMS AND ABBREVIATIONS

NA2, etc. = Reflects the other dose level(s) A+C	ABBREVIATION OR TERM	DEFINITION OR EXPLANATION
NAP = Nonactivation: Positive Control NA1 = Nonactivation: Test Compound Dos NA2, etc. = Reflects the other dose level(s) A+C = Negative Chemical Control for ACP A-C = Activation: Solvent Control ALI = Activation: Homogenate Control (ALI or A+T = Activation: Homogenate Control (ACP = Activation: Positive Control ACP = Activation: Positive Control ACT = Activation Test LI = Liver Tissue Activation Fraction LU = Lung Tissue Activation Fraction LU = Lung Tissue Activation Fraction Existence of Existence Activation Fraction Existence of Existence Activation Fraction TE = Testes Tissue Activation Fraction 1,2, etc. = Dose Levels CONCENTRATION All test compound dose levels are expressed as a whole number followed by an exponent (negative) identified by the appropriate units. Example: 0025-2PCT = 0,25 percent concentration POPU Total number of viable cells in the plating sample raised to some exponent printed directly below the abbreviation (i.e., EP + 6 = x 10 ⁶). MUT 1 Total number of mutants or convertants obtained from the sample plated raised to some exponent printed directly below the abbreviation (i.e., EP + 0 = 10 ⁰). For strain D4, MUT 1 represents the number of ADE+ convertants. MUT 2 Only used for strain D4 and represents the number of TRY+ convertants in the plated sample. FREQ 1 The calculated mutation or gene conversion frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value.	COMPOUND	
A-C ALI or A+T ALI or Activation: Homogenate Control (ACP ACT = Activation: Positive Control ACT = Activation Test LI = Liver Tissue Activation Fraction LU = Lung Tissue Activation Fraction KI = Kidney Tissue Activation Fraction Existe Tissue Activation Fraction TE = Testes Tissue Activation Fraction TE = Testes Tissue Activation Fraction Teste Tissue Activation Fraction Teste Tissue Activation Fraction Teste Tissue Activation Fraction To Dose Levels CONCENTRATION All test compound dose levels are expressed as a whole number followed by an exponent (negative) identified by the appropriate units. Example: 0025-2PCT = 0.25 percent concentration POPU Total number of viable cells in the plating sample raised to some exponent printed directly below the abbreviation (i.e., EP + 6 = x 10 ⁶). MUT 1 Total number of mutants or convertants obtained from the sample plated raised to some exponent printed directly below the abbreviation (i.e., EP + 0 = 10 ⁰). For strain D4, MUT 1 represents the number of TRY+ convertants in the plated sample. MUT 2 Only used for strain D4 and represents the number of TRY+ convertants in the plated sample. FREQ 1 The calculated mutation or gene conversion frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value. FREQ 2 Only used for strain D4 and represents the TRY+ conversion frequency.	TEST CODES	NAP = Nonactivation: Positive Control NAI = Nonactivation: Test Compound Dose 1
LU = Lung Tissue Activation Fraction KI = Kidney Tissue Activation Fraction TE = Testes Tissue Activation Fraction 1,2, etc. = Dose Levels CONCENTRATION All test compound dose levels are expressed as a whole number followed by an exponent (negative) identified by the appropriate units. Example: 0025-2PCT = 0.25 percent concentration POPU Total number of viable cells in the plating sample raised to some exponent printed directly below the abbreviation (i.e., EP + 6 = x 10 ⁶). MUT 1 Total number of mutants or convertants obtained from the sample plated raised to some exponent printed directly below the abbreviation (i.e., EP + 0 = 10 ⁰). For strain D4, MUT 1 represents the number of ADE+ convertants. MUT 2 Only used for strain D4 and represents the number of TRY+ convertants in the plated sample. FREQ 1 The calculated mutation or gene conversion frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value. FREQ 2 Only used for strain D4 and represents the TRY+ conversion frequency.		A-C = Activation: Solvent Control ALI
whole number followed by an exponent (negative) identified by the appropriate units. Example: 0025-2PCT = 0.25 percent concentration POPU Total number of viable cells in the plating sample raised to some exponent printed directly below the abbreviation (i.e., EP + 6 = x 10 ⁶). MUT 1 Total number of mutants or convertants obtained from the sample plated raised to some exponent printed directly below the abbreviation (i.e., EP + 0 = 10 ⁰). For strain D4, MUT 1 represents the number of ADE+ convertants. MUT 2 Only used for strain D4 and represents the number of TRY+ convertants in the plated sample. FREQ 1 The calculated mutation or gene conversion frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value. FREQ 2 Only used for strain D4 and represents the TRY+ conversion frequency.		LU = Lung Tissue Activation Fraction KI = Kidney Tissue Activation Fraction TE = Testes Tissue Activation Fraction
Total number of viable cells in the plating sample raised to some exponent printed directly below the abbreviation (i.e., EP + 6 = x 106). MUT 1 Total number of mutants or convertants obtained from the sample plated raised to some exponent printed directly below the abbreviation (i.e., EP + 0 = 100). For strain D4, MUT 1 represents the number of ADE+ convertants. MUT 2 Only used for strain D4 and represents the number of TRY+ convertants in the plated sample. FREQ 1 The calculated mutation or gene conversion frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value. FREQ 2 Only used for strain D4 and represents the TRY+ conversion frequency.	CONCENTRATION	whole number followed by an exponent (negative)
raised to some exponent printed directly below the abbreviation (i.e., EP + 6 = x 10 ⁶). MUT 1 Total number of mutants or convertants obtained from the sample plated raised to some exponent printed directly below the abbreviation (i.e., EP + 0 = 10 ⁰). For strain D4, MUT 1 represents the number of ADE+ convertants. MUT 2 Only used for strain D4 and represents the number of TRY+ convertants in the plated sample. FREQ 1 The calculated mutation or gene conversion frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value. FREQ 2 Only used for strain D4 and represents the TRY+ conversion frequency.		Example: 0025-2PCT = 0.25 percent concentration
from the sample plated raised to some exponent printed directly below the abbreviation (i.e., EP + 0 = 100). For strain D4, MUT 1 represents the number of ADE+ convertants. MUT 2 Only used for strain D4 and represents the number of TRY+ convertants in the plated sample. FREQ 1 The calculated mutation or gene conversion frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value. FREQ 2 Only used for strain D4 and represents the TRY+ conversion frequency.	POPU	raised to some exponent printed directly below the
FREQ 1 The calculated mutation or gene conversion frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value. FREQ 2 Only used for strain D4 and represents the TRY+ conversion frequency.	MUT 1	from the sample plated raised to some exponent printed directly below the abbreviation (i.e., $EP + 0 = 10^{0}$). For strain D4, MUT 1 represents the
frequency times the negative exponent written directly below. For strain D4, FREQ 1 represents the ADE+ value. FREQ 2 Only used for strain D4 and represents the TRY+ conversion frequency.	MUT 2	Only used for strain D4 and represents the number of TRY+ convertants in the plated sample.
conversion frequency.	FREQ 1	frequency times the negative exponent written directly below. For strain D4, FREQ 1
CONTAM Presence of contamination on any plates	FREQ 2	Only used for strain D4 and represents the TRY+ conversion frequency.
tresence of concamination on any places.	CONTAM	Presence of contamination on any plates.



DATA TABLE TERMS AND ABBREVIATIONS (continued)

ABBREVIATION OR TERM	DEFINITION OR EXPLANATION
AAF	2-Acetylaminofluorene
DMSO	Dimethylsulfoxide
DMN	Dimethylnitrosamine
EMS	Ethylmethanesulfonate
QM	Quinacrine Mustard
NÉ	Nitrofluorene
ANTH	2-Amino Anthracene
AMQ	8-Amino Quinoline
SPECIES	Animal Strains
SPRDAW	Sprague Dawley Rats
ICRFLO	Flow ICR Random Bred Mice
RHESUS	Rhesus Monkey (<u>Macaca mulatta</u>)
MIXEDB	Dog, Mixed Breed
NEWZEA	New Zealand White Rabbit
UG	Microgram
UM	Micromole
ADE	Adenine
TRY	Tryptophan



V. <u>INTERPRETATION OF RESULTS AND CONCLUSIONS</u>

The test compound, FDA 75-88, Niacin (nicotinic acid), was evaluated for genetic activity in a series of in vitro microbial assays with and without metabolic activation. The following results were obtained:

- A. Salmonella typhimurium
- Plate tests

The results of these tests were negative.

2. Nonactivation suspension tests

The results of these tests were negative.

3. Activation suspension tests

The results of these tests were negative. The following test was repeated with TA-1537 using mouse lung tissue.

This test was repeated for the following reason: In the initial test this strain exhibited increased revertant frequency at all three doses. The repeat test was negative.

- B. <u>Saccharomyces cerevisiae</u>
- 1. Nonactivation suspension tests

The results of these test were negative.

2. Activation suspension tests

The results of these tests were negative.

C. Conclusions

The test compound, FDA 75-88, Niacin (nicotinic acid), did not exhibt mutagenic activity in any of the assays employed in these studies.

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Robert J. Weir, Ph.D. D. Vice President

VI. <u>EXPLANATION OF EVALUATION PROCEDURES FOR PLATE ASSAYS</u>

Plate test data consist of direct revertant colony counts obtained from a set of selective agar plates seeded with populations of mutant cells suspended in a semisolid overlay. Because the test chemical and cells are incubated in the overlay for 2-3 days, and a few cell divisions occur during the incubation period, the test is semiquantitative in nature. Although these features of the assay reduce the quantitation of results, they provide certain advantages not contained in a quantitative suspension test.

- The small number of cell divisions permits potential mutagnes to act on replicating DNA which is often more sensitive than non-replicating DNA.
- The combined incubation of the compound and the cells in the overlay permit constant exposure of the indicator cells for 2-3 days.

A. Surviving Populations

Plate test procedures do not permit exact quantitation of the number of cells surviving chemical treatment. At low concentrations of the test chemical, the surviving population on the treatment plates is essentially the same as the negative control plate. At high concentrations, the surviving population is usually reduced by some fraction. Our protocol normally employs dose levels that are selected such that the highest dose will show slight toxicity (as determined by subjective criteria) and several doses ranging down 1 to 2 logs lower.

B. Dose Response Phenomena

The demonstration of dose-related increases in mutant counts is an important criterion in establishing mutagenicity. Factors which may modify dose response results for a mutagen would be the selection of doses that are too low (usually mutagenicity and toxicity are related). If the highest dose is far lower than a toxic concentration, no increases may be observed over the dose range selected. Conversely, if the lowest dose employed is highly cytotoxic, the test chemical may kill any mutants that are induced and the compound will not appear to be mutagenic.

C. Control Tests

Positive and negative control assays are conducted with each experiment and consist of direct acting mutagens for nonactivation assays and mutagens that require metabolic biotransformation in activation assays. Negative controls consist of the test compound solvent in the overlay agar with the other essential components. The negative control plate for each strain gives a reference point to which the test data are compared. The positive control assay is conducted to demonstrate that the test systems are functional with known mutagens.



D. <u>Evaluation Criteria for Ames Assay</u>

Because the procedures used to evaluate the mutagenicity of the test chemical are semiquantitative, the criteria used to determine positive effects are inherently subjective and are based primarily on a historical data base. Most data sets are evaluated using the following criteria:

1. Strains TA-1535, TA-1537, and TA-1538

If the solvent control value is within the normal range, a chemical that produces a positive dose response over three concentrations with the lowest increase equal to twice the solvent control value is considered to be mutagenic.

2. Strains TA-98, TA-100, and D4

If the solvent control value is within the normal range, a chemical that produces a positive dose response over three concentrations with the highest increase equal to twice the solvent control value for TA-100 and two to three times the solvent control value for strains TA-98 and D4 is considered to be mutagenic. For these strains, the dose response increase should start at approximately the solvent control value.

Pattern

Because TA-1535 and TA-100 were both derived from the same parental strain (G-46) and because TA-1538 and TA-98 were both derived from the same parental strain (D3052), there is a built-in redundancy in the microbial assay. In general the two strains of a set respond to the same mutagen and such a pattern is sought. It is also anticipated that if a given strain, e.g. TA-1537, responds to a mutagen in nonactivation tests it will generally do so in activation tests. (The converse of this relationship is not expected.) While similar response patterns are not required for all mutagens, they can be used to enhance the reliability of an evaluation decision.

4. Reproducibility

If a chemical produces a response in a single test that cannot be reproduced in one or more additional runs, the initial positive test data loses significance.

The preceding criteria are not absolute and other extenuating factors may enter into a final evaluation decision. However, these criteria are applied to the majority of situations and are presented to aid those individuals not familiar with this procedure. As the data base is increased, the criteria for evaluation can be more firmly established.



VII. EXPLANATION OF EVALUATION PROCEDURES FOR SUSPENSION ASSAYS

Data obtained from mutagenicity tests are evaluated on a test by test basis followed by an examination of the total response pattern using all the data. To facilitate this type of evaluation, we have prepared two separate formats in which data are processed. The first is the Compound Summary Backup Detail Sheet, which details the essential raw data from each experiment showing surviving population counts, total mutant or convertant counts, as well as, calculated mutation frequencies. This format permits close examination of each set of test data. The following considerations are part of any assessment.

A. Surviving Population Counts

A certain level of chemically-induced toxicity is anticipated, but occasionally isolated tests or groups of tests show very low (<25%) survival compared to the tissue controls. Such isolated decreases may result from improper dilution procedures or defective growth media and decrease confidence in the calculated mutation frequencies especially if the total mutant counts appear unaffected. Data of this type are generally unacceptable and these experiments are routinely repeated at a lower dose level to reduce killing and increase confidence in the nature of the response.

B. Total Mutant Counts

For nonmutagens, the mutant/surviving population ratio should be roughly equivalent for each test point in a given experiment. If the cell number drops in response to killing, the mutant number should decrease proportionately. A mutagenic chemical, however, will produce an altered mutant/surviving population ratio. Mutant numbers as well as calculated frequencies are compared to the negative control data. In certain instances, the mutant frequencies will increase with little or no change in the absolute number of mutants especially where the test chemical is toxic. Data of this type, although not necessarily aberrant, or even rare, must be viewed with special care to ensure that the increased frequencies were not the result of selective toxicity of the test chemical for the his cells. This phenomenon, referred to as selection, can lead to erroneous conclusions. Thus we attempt to keep the surviving population of cells high and look for positive responses that show increases in both numbers of mutants and mutation frequencies. Again, occasional isolated fluctuations in mutant counts are found that can be attributed to improper pipetting or media contamination. These fluctuations are usually easy to identify by inspection of the other data points in the experiment which will be negative.



C. Dose Response Phenomena

Dose-related increases in mutants and mutation frequencies are the most convincing data to have in assessing mutagenic activity of chemicals. In some cases, however, dose-related increases are not observed for mutagens. This depends considerably on the dose levels selected. The figure on the following page illustrates how one might obtain various types of dose-related responses by a mutagen based solely on dose selection. It also emphasizes the need to keep dose levels within a relatively low range of toxicity so that data are consistently on the uphill side of the hypothetical curve.

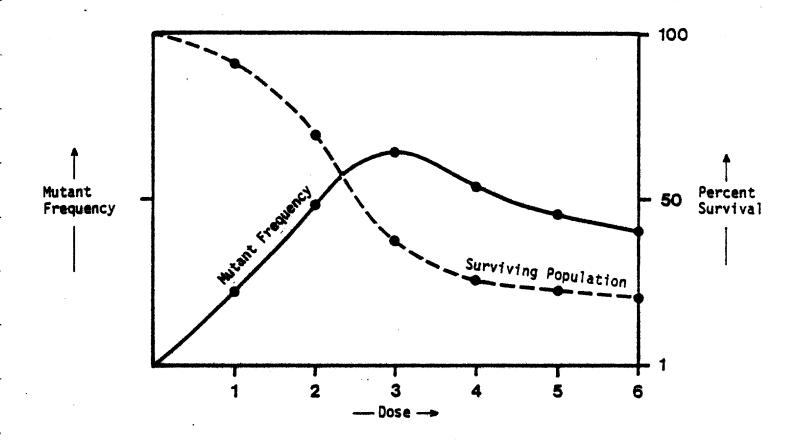
D. Control Tests

Positive and negative control tests are conducted with each experiment and consist of direct acting positive agents for nonactivation assays and chemicals that require metabolic transformation for activation assays. In nonactivation assays, the NAN control contain the test chemical solvent plus cells, but no chemical, and is used as a reference to assess the level of response obtained in the various tests. It is not possible at this time to put precise cut-off points where negative responses become positive responses. A statistical component for our computer program is under development and will be included when available. Positive controls are only used as relative reference points and to demonstrate that the system is functioning with known mutagens. In activation assays, three types of negative controls are run: (1) A solvent control minus the chemical and minus the activation system (A-C); (2) a control plus the positive control chemical minus the activation system (A+C); and (3) a control containing the activation system and the test chemical solvent (ALI or ALU). All three controls are used collectively to assess the level of response in the various activation tests. A chemical may appear positive when compared to an A-C control but not when compared to an A+T control. The value of each of the above controls with respect to their weight in evaluation is ALI or ALU > A-C > A+C.

The other data format is the Compound Frequency Summary Report sheet in which all the calculated frequencies obtained for a given compound are displayed in a table. This format permits an overview of all data. The points form a matrix of information that should present a consistent pattern. Nonmutagens should produce a matrix with data frequencies clustered around the negative control values. Occasional random high or low fluctuations are not uncommon and seldom indicate true genetic activity. Mutagenic chemicals should, on the other hand, produce a set of consistent responses that demonstrate a logical pattern. The patterns depend on the mutagenic specificity of the chemical but can be easily recognized in the Compound Frequency Summary Report format.

These mutagenicity assays are designed to optimize the probability of recognizing mutagens from nonmutagens and, in most cases, they work well. Occasionally, the data points are such that a definitive conclusion cannot be made without additional data.





HYPOTHETICAL EXPERIMENT

- (1) Dose levels 1,2 & 3 were used
- (2) Dose levels
 2, 3 & 4 were used
- (3) Dose levels
 3, 4 & 5 were used

OBSERVED DOSE RESPONSE

A typical positive dose response set of data would be obtained.

The intermediate dose level shows a higher mutation frequency than both the low dose and the high dose.

Here an inverted dose response would be observed with the highest dose level showing the lowest response.

APPENDIX Tabulation of Data



EXPERIMENT			223-76-2102 DETECTOR TA100	SPE	CIES	PROJECT 2672	DATE - 07/22/77
COMPOUND	TEST	ORG ID	CONCENTRATION	POPU EP+6	MUTI	FREGI EP-8	CONTAN
CORPORE	MAN	10	SOLVENT	0252		85.71	CONTAN
	NAP		EMS 0.066%	0616	5547	900.49	•
000059676	NAI		0016-3 PCT.	0668	8473	70.81	0
000059676	SAM		9008-3 PCT.	0803	0435	54.17	•
000059676	EAN.		0004-3 PCT.	0636	9468	73.58	•

EXPERIMENT			223-76-2102 DETECTOR TA1535	SPECIES		PROJECT 2672	DATE - 07/22/77
COMPOUND	TEST	ORG ID	CONCENTRATION	POPU EP+6	MUT1 EP+0	FREQ1 EP-8	CONTAH
	NAN		SOLVENT	1310	0019	1.45	•
	NAP		EHS 0.2%	0852	5840	. 685.45	. 8
000059676	NA1		0016-3 PCT.	0979	0035	3.50	•
000059676	NAZ		0006-3 PCT.	0868	0021	2.42	0
000059676	NA3		0004-3 PCT.	0893	0028	3.14	0

EXPERIMENT	CONTRACT 710101	223-76-2102 DETECTOR TA1537	SPECIES	PROJECT 2672	DATE - 07/22/77
COMPOUND	ORG TEST 1D	CONCENTRATION	POPU MUTI EP+6 EP+0	FREQ1 EP-8	CONTAN
	NAN	SOLVENT	0466 0054	11.59	0
	NAP	QH 13 UG/ML	0235 0553	235.32	0
000059676	NA1	0016-3 PCT.	4595 0841	6.89	
000059676	NAZ	0008-3 PCT.	1462 0042	2.67	• .
000059676	EAM	0004-3 PCT.	0350 0034	9.71	•

EXPERIMENT			223-76-2102 DETECTOR TA1538	SPECIES		PROJECT 2672	DATE - 07/22/77
COMPOUND	TEST	1D 086	CONCENTRATION	POPU EP+6	HUT1 EP+0	FREQ1 EP-8	CONTAN
	NAN		SOLVENT	0403	0022	5.46	•
	HAP		NF 667 UG/HL	0376	9614	163.30	•
000059676	NAI		0016-3 PCT.	0512	0018	3.52	0
000059676	NAZ		0008-3 PCT.	8414	0016	3.86	•
000059676	EAM		0004-3 PCT.	046.1	0011	2.39	•

EXPERIMENT		223-76-2102 Detector Ta98	SPECIES	PROJECT 2672	DATE - 67/22/77
COMPOUND	OR6 TEST ID	CONCENTRATION	POPU MUT1 EP+6 EP+0	FREQ1 EP-8	CONTAM
	NAN	SOLVENT	6962 0133	13.83	•
	NAP	NF 667 UG/ML	0834 0598	71.70	•
000059676	NA1	0016-3 PCT.	1507 0156	10.35	•
008059676	NAZ	0008-3 PCT.	1344 0103	7.66	•
000059676	EAN	0004-3 PCT.	1238 9194	8.40	•

	CON	ITRACT	223-76	-5165						
EXPERIMENT	7109	902	DETECTOR 8000D4		SPECIES		· /			DATE - 07/22/77
		0R6			POPU	MUTI	MUT2	FREGI	FREQZ	
COMPOUND	TEST	10	CONCEN	TRATION	EP+4	EP+1	EP+1	EP-5	EP-5	CONTAN
	NAN		SOLVEN	T	1175	0234	0031	19.91	2.64	1
	NAP		EMS 1.	0 %	1463	1600	1142	109.36	78.06	•
000059676	NAI		0033-2	PCT.	1386	0258	0088	18.61	.063	. 0
000059676	NA2		0165-3	PCT.	1254	0199	0060	15.87	4.78	.0
000059676	EAM.		0825-4	PCT.	1519	0291	0081	19.16	5.33	•

EXPERIMENT	CONTRACT 710301		223-76-2102 DETECTOR TA100	SPE	CIES IC	PROJECT 2672 RFLO/MOUSE	DATE - 07/22/77
COMPOUND	TEST	ORG ID	CONCENTRATION	POPU EP+6	MUT1 EP+0	FREGI EP-8	CONTAH
	A+C		DMN 98 UH/HL	•98 9	0835	84.43	•
	A-C		SOLVENT	0899	0502	55.84	•
	ALI		TISSUE	1545	0867	57.41	•
	ALU		TISSUE	1869	1097	58.69	•
	ACP	LI	DMN 98 UM/ML	1160	2112	182.07	•
	ACP	LU	DHN 90 UM/ML	0496	0456	91.94	•
000059676	ACT	LII	0016-3 PCT.	0673	0447	66.42	0
000059676	ACT	LIZ	0008-3 PCT.	0612	0454	74.18	•
000059676	ACT	LI3	6064-3 PCT.	0528	0362	68.56	•
000059676	ACT	FOI	0016-3 PCT.	1265	0319	24.82	•
000059676	ACT	FnS	0008-3 PCT.	1321	0375	28.39	•
00059676	ACT	LÚ3	0004-3 PCT.	1534	0461	34.05	

		CONTRACT		223-76-2102			PROJECT 2672	
EXPERIMENT		710303		DETECTOR TA1535	SPECIES		ICRFLO/MOUSE	DATE - 07/22/77
			0 86		POPU	MUTI		
	COMPOUND	TEST	10	CONCENTRATION	EP+6	EP+0	EP-8	CONTAM
		A+C		DHN 98 UH/HL	9649	0946	7.09	•
		A-C		SOLVENT	0431	0016	4.18	•
		ALI		TISSUE	0273	0016	5.86	•
		ALU		TISSUE	9346	0026	8.09	•
		ACP	LI	DHN 90 UN/ML	0687	0539	78.46	•
		ACP	LU	DHN 98 UM/HL	0570	0052	9.12	0
	000059676	ACT	LII	6616-3 PCT.	8489	0021	5.73	0
	000059676	ACT	F15	0008-3 PCT.	9496	001	3.20	•
	000059676	ACT	LI3	0004-3 PCT.	9754	0020	2.65	0
	000059676	ACT	LUI	0016-3 PCT.	0708	002	3.53	0
	000059676	ACT	LU2	0008-3 PCT.	0670	002	3.28	•
	088059676	ACT	LU3	0004-3 PCT.	8754	002	3.71	0

	CONTRACT		223-76-2102			PROJECT 2672	
EXPERIMENT	710802		DETECTOR TA1537	SPE	CIES	ICRFLO/HOUSE	DATE - 07/22/7
COMPOUND	TEST	ID ORG	CONCENTRATION	POPU EP+6	HUT1 EP+0		CONTAM .
	A+C		AHQ 333 UG/HL	2235	0208	9.31	•
	A-C		SOLVENT	2639	0166	6.29	•
	ALI		TISSUE	0769	0098	12.74	•
	ALU		TISSUE	0768	9854	7.03	•
	ACP	LI	ANQ 333 UG/HL	2337	2282	97.65	•
	ACP	LU	ANG 333 UG/ML	9392	9016	4.19	2
000059676	ACT	LH	0016-3 PCT.	0489	0119	24.34	•
000059676	ACT	L12	0008-3 PCT.	1211	0124	10.24	0
000059676	ACT	LI3	0004-3 PCT.	1415	0080	5.65	0
000059676	ACT	FnJ	0016-3 PCT.	0358	0169	47.21	•
000059676	ACT	Fn5	9008-3 PCT.	1023	0182	17.79	G
000059676	ACT	LU3	0004-3 PCT.	0645	0172	26.67	0

	CON	ITRACT	223-76-2102			PROJECT 2672	
EXPERIMENT	7112	101	DETECTOR TA1538	SPE	CIES	ICRFLO/HOUSE	DATE - 07/22/77
		0Ř6		POPU	MUTI	7 7-	
COMPOUND	TEST	10	CONCENTRATION	EP+6	EP+0	EP-8	CONTAM
	A+C		ANTH 67 UG/ML	1219	0115	9.43	•
	A-C		SOLVENT	1226	0104	8.48	. 1
	ALI		TISSUE	1143	0063	5.51	• •
	ALU		TISSUE	1145	0085	7.42	1
	ACP	LI	ANTH 67 UG/ML	1117	1667	149.24	1
	ACP	LU	ANTH 67 UG/ML	0302	0230	76.16	. 1
000059676	ACT	LH	0016-3 PCT.	1186	0103	8.67	2
000059676	ACT	LIZ	0008-3 PCT.	1759	0125	7.11	•
000059676	ACT	LI3	0004-3 PCT.	1324	0107	8.08	0
000059676	ACT	Fol	0016-3 PCT.	1279	0122		•
000059676	ACT	F05	0008-3 PCT.	1436	0092	6.41	1
000059676	ACT	LU3	0004-3 PCT.	1245	0115	9.24	0

EXPERIMENT		NTRACT 304	223-76-2162 DETECTOR TA98	SPE	CIES ICRI	DATE - 07/22/77	
COMPOUND	TEST	10 086	CONCENTRATION	PGPU EP+6	MUT1 EP+0	FREQ1 EP-8	CONTAH
	A+C		ANTH 67 UG/ML	1797	0170	9.46	•
	A-C		SOLVENT	1513	0133	8.79	0
	ALI		TISSUE	0810	0174	21.48	0
	ALU		TISSUE	1095	0161	14.70	•
	ACP	LI	ANTH 67 UG/ML	9629	0549	87.28	•
	ACP	LU	ANTH 67 UG/HL	1134	0846	74.60	0
000059676	ACT	LII	0016-3 PCT.	0730	0197	26.99	0
000059676	ACT	LIS	0008-3 PCT.	9841	0155	18.43	0
000059676	ACT	LI3	0004-3 PCT.	0952	0143	15.02	0
000059676	ACT	Fn1	0016-3 PCT.	1059	0177	16.71	0
000059676	ACT	LU2	0008-3 PCT.	1050	0145	13.41	0
000059676	ACT	LU3	9994-3 PCT.	1168	0145	12.41	•

EXPERIMENT	CONTRACT 712902		223-76-2102 Detector 400004	SPE	CIES	PRO ICRFLO/	DATE - 07/22/77		
COMPOUND	TEST	ORG ID	CONCENTRATION	POPU EP+4	HUT1 EP+1		FREQ1 EP-5	FREQ2 EP-5	CONTAN
	A+C		DMN 90 UM/HL	1184	0082	-	6.93	15.63	0
	A-C		SOLVENT	1072	0074	0243	6.90	22.67	• .
	ALI		TISSUE	1474	0153	0150	10.36	8.14	•
	ALU		TISSUE	1320	0068	0198	5.15	15.00	•
	ACP	LI	DHN 98 UH/HL .	1555	9652	1122	53.36	91.82	0
	ACP	LU	DMN 90 UM/ML	1061	0202	0184	19.04	17.34	•
000059676	ACT	LİI	0033-2 PCT.	1767	0023	0012	1.30	0.68	0
000059676	ACT	LI2	0165-3 PCT.	2083	0025	0612	1.20	0.58	•
000059676	ACT	£13	0825-4 PCT.	1,166	9966	0020	5.66	1.72	•
000059676	ACT	rul	0033-2 PCT.	1514	0028	0017	1.85	1.12	•
000059676	ACT	FnS	0165-3 PCT.	1827	0029	0022	1.59	1.20	•
000059676	ACT	LU3	0825-4 PCT.	1522	0996	0085	6.31	6 60	

	CON	ITRACT	223-76-2102				
EXPERIMEN	T 7110	03	DETECTOR TALOS	SPE	CIES SPR	DAW/RAT	DATE - 07/22/77
		OR6		POPU	MUTI	FAEQ1	
COMPOUND	TEST	10	CONCENTRATION	EP+6	6+43	EP-8	CONTAH
	A+C		DHN 90 UH/HL	8484	0084	20.79	•
	A-C		SOLVENT	8499	0093	18.64	•
	ALI		TISSUE	0230	8192	83.48	•
	ALU		TISSUE	8487	0315	64.68	•
	ACP	LI	DHN 90 UH/HL	0319	0791	247.96	•
	ACP	LU	DHN 90 UH/HL	8779	0410	52.63	•
000059676	ACT	LII	0016-3 PCT.	9446	0073	16.37	•
000059676	ACT	L12	9008-3 PCT.	0205	0064	31.22	•
000059676	ACT	LI3	0004-3 PCT.	0470	0045	9.57	•
000059676	ACT	FOI	0016-3 PCT.	0493	0263	53.35	0
660659676	ACT	LU2	0008-3 PCT.	0824	0379	46.88	•
000059676	ACT	LU3	0004-3 PCT.	0951	0438	46.06	

	CON	TRACT	223-76-2102				PROJECT 2672	
EXPERIMENT	7111	03	DETECTOR	TA1535	SPE	CIES	SPRDAW/RAT	DATE - 07/22/77
		ORG			POPU	HUTI	FRE41	
COMPOUND	TEST	10	CONCENTRA	TION	EP+6	EP+0	EP-8	CONTAN
	A+C		DHN 98 U	4/ML	1386	0019	1.37	• ,
	A-C		SOLVENT		0841	6032	3.60	•
	ALI		TISSUE		1442	0038	2.64	•
	ALU		TISSUE		1420	0025	1.76	0
	ACP	LI	DMN 90 U	H/HL	1141	4694	411.39	0
	ACP	LU	DHN 90 U	H/HL	1393	0018	1.29	•
000059676	ACT	LII	0016-3 P	CT.	1330	8844	3.31	•
000059676	ACT	LI2	0008-3 P	CT.	1264	0037	2.93	•
000059676	ACT	LI3	0004-3 P	cT.	8946	8844	4.65	0
00059676	ACT	LUI	0016-3 P	CT.	1180	0021	1.78	•
000059676	ACT	Fn5	9908-3 P	ct.	1362	0027	1.98	
000059676	ACT	Fn3	0004-3 R	CT.	1142	0025	2.19	•

EXPERIMENT	CON 7136	ITRACT 153	223-76-2102 DETECTOR TA1537	. SPE	CIES SPE	PROJECT 2672 RDAW/RAT	DATE - 07/22/77
		ORG					DAIL 01/22/11
COMPOUND	TEST	10	CONCENTRATION	POPU EP+6	HUT1 EP+0	FREU) EP-8	CONTAN
	A+C		AHQ 333 UG/HL	9658	0014	2.13	0
	A-C		SOLVENT	0692	0043	6.21	•
	ALI		TISSUE	0496	0036	7.26	•
	ALU		TISSUE	0687	0018	2.62	•
	ACP	LI	AMQ 333 UG/ML	0437	0500	116.25	•
	ACP	LU	AMQ 333 UG/HL	0696	0022	3.16	•
000059676	ACT	LII	00163 PCT.	0392	0030	7,65	•
000059676	ACT	F15	00083 PCT.	0371	0030	8.09	•
000059676	ACT	LI3	0004-3 PCT.	0525	0040	7.62	•
000059676	ACT	LUI	00163 PCT.	0514	0036	7.00	•
000059676	ACT	LU2	00083 PCT.	0602	0041	6.81	•
000059676	ACT	LU3	0004-3 PCT.	0801	0034	4.24	·

EXPERIMENT		TRACT	223-76-2102 DETECTOR TA1538	SPE	CIES S	PROJECT 2672 PRDAW/RAT	DATE - 07/22/77
COMPOUND	TEST	ORG ID	CONCENTRATION	POPU EP+6	MUT1 EP+0	FREQ1 EP-8	CONTAM
	A+C		ANTH 67 UG/ML	0956	0101	10.56	•
	A-C		SOLVENT	1244	0063	5.06	2
	ALI		TISSUE	0536	0058	10.82	•
	ALU		TISSUE	0552	0057	10.33	•
	ACP	LI	ANTH 67 UG/ML	0977	0869	88.95	٥
	ACP	LU	ANTH 67 UG/ML	0443	0709	160.05	6
000059676	ACT	LII	0016-3 PCT.	0441	9988	19.95	2
000459676	ACT	L12	0008-3 PCT.	0822	0052	6.33	2
000059676	ACT	LI3	9004-3 PCT.	0641	0029	4.52	2
# 00059676	ACT	FD)	0016-3 PCT.	0416	0045	10.77	•
000059676	ACT	rn5	6008-3 PCT.	0480	8628	7.00	•
000059676	ACT	LU3	0004-3 PCT.	0518	0033	6.37	0

EXPERIMEN		TRACT	223-76-2102 DETECTOR TA98	SPE	CIES SPE	DATE - 07/22/77	
COMPOUND	TEST	10 086	CONCENTRATION	POPU EP+6	MUT1 EP+0	FREU1 EP-8	CONTAH
	A+C		ANTH 67 UG/HL	8915	0120	13.11	•
	A-C		SOLVENT	8495	9963	15.56	•
	ALI		TISSUE	0455	0166	36.48	•
	ALU		TISSUE	1127	0201	17.63	•
	ACP	LI	ANTH 67 UG/ML	0483	1435	297.10	•
	ACP	LU	ANTH 67 US/HL	0961	1200	124.67	. •
000059676	ACT	LII	0016-3 PCT.	0543	0193	35.54	•
000059676	ACT	F15	0008-3 PCT.	0702	0224	31.91	•
000059676	ACT	£I3	0004-3 PCT.	0386	0164	42.49	0
000059676	ACT	rai	0016-3 PCT.	0608	0211	34.70	•
000059676	ACT	Fn5	0008-3 PCT.	9908	0171	16.63	•
000059676	ACT	LU3	0004-3 PCT.	0811	0183	22.56	۵

	CON	TRACT	223-76-2102			PRO	12		
EXPERIMENT	7129	01	DETECTOR 0000D4	SPE	CIES S	PRDAW	RAT		DATE - 07/22/77
		086		POPU	MUTI	MUTZ	FREQL	FREQ2	
COMPOUND	TEST	10	CONCENTRATION	EP+4	EP+1	EP+1	EP-5	EP-5	CONTAN
	A+C		DHN 90 UH/ML	1051	0164	0107	15.60	10.18	•
	A-C		SOLVENT	1533	0197	0114	12.85	7.44	•
	ALI		TISSUE	1306	0196	0135	15.01	10.34	•
	ALU		TISSUE	1091	0171	0112	15.67	10.27	•
	ACP	LI	DHN 98 UH/ML	1173	1294	0840	110.32	71.61	•
	ACP	LU	DHN 90 UM/HL	1118	0194	0086	17.35	7.69	0
000059676	ACT	LII	0033-2 PCT.	8967	6109	0121	11.27	12.51	•
004059676	ACT	LIS	0165-3 PCT.	0961	0142	0112	14.78	11.65	•
000059676	ACT	L13	0825-4 PCT.	1066	0138	0056	12.95	5.25	•
000059676	ACT	LU1	0033-2 PCT.	1275	0133	0049	10.43	3.84	0
000059676	ACT	LUZ	0165-3 PCT.	1227	0104	0053	8.48	4.32	0
999859676	ACT	LU3	0825-4 PCT.	1253	0145	0060	11.57	4.79	

CONTRAC EXPERIMENT 710401			223-76-2102 DETECTOR TA160	SPE	CIES HHE	DATE - 07/22/77	
		086		POPU	MUT1	FREGI	
COMPOUND	TEST	ID	CONCENTRATION	EP+6	EP+0	EP-8	CONTAH
	A+C		DHN 90 UH/HL	0839	0737	87.84	•
	A-C		SOLVENT	0863	0 557	64.54	•
	ALI		TISSUE	0874	9790	80.09	
	ALU		TISSUE	0823	8 578	74.23	6
	ACP	LI	DHN 90 UM/ML	9617	1756	284.60	e
	ACP	LU	DMN 98 UM/HL	1013	9701	69.20	•
000059676	ACT	LII	0016-3 PCT.	1142	0739	64.71	0
000059676	ACT	LIS	0008-3 PCT.	1076	0747	69.81	0
000059676	ACT	LI3	0004-3 PCT.	1049	9654	62.35	0
000059676	ACT	LUI	0016-3 PCF.	8948	0715	75.42	0
000059676	ACT	LU2	9898-3 PCT.	1027	9639	62.22	0
000059676	ACT	LU3	0004-3 PCT.	0937	0805	85.91	•

EXPERIMENT		TRACT 02	223-76-2102 DETECTOR TA1535	SPE	CIES	PROJECT 2672 RHESUS/MONKEY	DATE - 07/22/77
COMPOUND	TEST	OKG ID	CONCENTRATION	POPU EP+6	NUT I	-	CONTAH
	A+C		DMN 96 UH/HL	9887	0050	5.64	•
	A-C		SOLVENT	8945	8026	2.96	0
	ALI		TISSUE	0903	0050	5.54	•
	ALU		TISSUE	0846	0032	3.78	•
	ACP	LI	DHN 90 UH/HL	1160	9666	56.98	•
	ACP	Lu	DMN 90 UH/HL	1028	0049	4.38	0
000059676	ACT	LII	0016-3 PCT.	1204	0066	5.30	• •
000059676	ACT	L12	0008-3 PCT.	1093	0069	6.31	0
990959676	ACT	LI3	0004-3 PCT.	1200	0046	4,00	
000059676	ACT	LUI	0016-3 PCT.	0841	0037	4.40	•
000059676	ACT	LU2	0008-3 PCT.	1138	0046	4.04	
000059676	ACT	LU3	8884-3 PCT.	1343	0055	4.10	•

EXPERIMENT			223-76-2102 DETECTOR TA1537	SPE	CIES RHE	DATE - 47/22/77	
		ORG		POPU	MUT1	FRE01	
COMPOUND	TEST	10	CONCENTRATION	EP+6	EP+0	EP-8	CONTAM
	A+C		AMQ 333 UG/ML	0352	0077	21.86	0
	A-C		SOLVENT	0641	9908	1.25	•
	ALI		TISSUE	0623	0031	4.98	•
	ALU	٠	TISSUE	1229	0090	7.32	•
	ACP	LI	AMQ 333 UG/ML	1134	0428	37.74	0
	ACP	LU	AMQ 333 UG/HL	1158	0158	13.64	•
000059676	ACT	LII	0016-3 PCT.	0733	0047	6.41	0
000059676	·ACT	FIS	0008-3 PCT.	0645	0031	4.81	0
000059676	ACT	LI3	0004-3 PCT.	0860	0019	2.21	•
000059676	ACT	LU1	0016-3 PCT.	1190	0053	4.45	0
004059676	ACT	Fn5	0008-3 °CT.	1084	0050	4.61	0
000059676	ACT	LU3	0004-3 °CT.	1960	9840	3.77	•

HEPORT EXR33 LITTON BIONETICS MUTAGENIC ACTIVITY SYSTEM COMPOUND SUMMARY BACKUP DETAIL

EXPERIMENT			223-76-2102 DETECTOR TA1538	SPE	CIES	PROJECT 2672 RHESUS/MONKEY	DATE - 07/22/77
COMPOUND	TEST	ORG ID	CONCENTRATION	POPU EP+6	MUT1 EP+0	FREQ1 EP-8	CONTAN
	A+C		ANTH 67 UG/ML	0700	0097	13.86	2
	A-C		SOLVENT	4689	0079	11.47	2
	ALI		TISSUE	0590	0073	12.37	. 2
	ALU		TISSUE	0002	9981	10.10	2
	ACP	LI	ANTH 67 UG/ML	6798	1229	154.01	2
	ACP	LU	ANTH 67 UG/ML	1046	0081	7.74	2
000059676	ACT	LII	0016-3; PCT.	9537	0112	20.86	2
000059676	ACT	F15	0008-3 PCT.	0677	0087	12.85	. 2
000059676	ACT	LI3	0004-3 PCT.	0826	0121	14.65	2
0,00059676	ACT	Lu1	0016-3 PCT.	0430	0094	21.86	2
000059676	ACT	LU2	0008-3 PCT.	0857	0082	9.57	2
000059676	ACT	LU3	0004-3 PCT.	0474	0066	13.92	2

REPORT EXR33 LITTON BIONETICS MUTAGENIC ACTIVITY SYSTEM COMPOUND SUMMARY BACKUP DETAIL

EXPERIMENT	CONTRACT 710403		223-76-2102 DETECTOR TA96	SPE	CIES RHE	DATE - 07/22/77	
	TEST	086 ID	CONCENTO A TION	POPU	MUTI	FRE41	
COMPOUND	1531	10	CONCENTRATION	EP+6	EP+0	EP-8	CONTAN
	A+C		ANTH 67 UG/HL	1452	0139	7.51	•
	A-C		SOLVENT	1660	0136	8.19	•
	ALI		TISSUE	0852	0124	14.55	• .
	ALU		TISSUE	1167	4122	10.45	
	ACP	LI	ANTH 67 UG/ML	1150	2319	201.65	0
	ACP	LU	ANTH 67 UG/ML	1789	0147	8.22	0
000059676	ACT	LII	9016-3 PCT.	1381	0175	12.67	0
000059676	ACT	L15	0008-3 PCT.	1213	0154	12.70	0
000059676	ACT	LI3	8884-3 PCT.	1203	0149	12.39	0
000059676	ACT	LU]	0016-3 PCT.	9669	0117	17.73	0
000059676	ACT	LU2	0008-3 PCT.	1035	0120	11.59	•
000059676	ACT	LU3	0004-3 PCT.	1195	0138	11.55	•

	CONTRACT		223-76-2102			PRO.	72		
EXPERIMENT	7113	02	DETECTOR 000004	SPE	CIES I	HESUS/	HONKEY		DATE - 07/22/77
		0H6		POPU	MUTI	MUT2	FREGI	FREQ2	
COMPOUND	TEST	ID	CONCENTRATION	EP+4	EP+1	EP+1	EP-5	EP-5	CONTAH
	A+C		DHN 90 UH/ML	1110	0244	0121	21.98	10.90	•
	A-C		SOLVENT	1343	0108	0098	8.04	7.30	0
	ALI		TISSUE	1385	0225	0086	16.25	6.21	0
	ALU		TISSUE	1268	9263	0098	20.74	7.73	•
	ACP	LI	DMN 90 UM/ML	1370	0935	0743	68.25	54.23	•
	ACP	LU	DHN 90 UM/ML	1515	0248	0089	20.46	7.34	1
000059676	ACT	LII	0033-2 PCT.	0958	0039	6018	4.07	1.00	•
000059676	ACT	FIS	0165-3 PCT.	0984	9048	0017	4.88	1.73	•
000059676	ACT	LI3	0825-4 PCT.	1052	6652	0023	4.94	2.19	1
000059676	ACT	LU1	0033-2 PCT.	1544	9966	0837	4.27	2.48	1
000059676	ACT	LU2	0165-3 PCT.	1053	0076	0024	7.22	2.28	0
000059676	ACT	LU3	0825-4 PCT.	1435	0102	0038	7.11	2.65	6